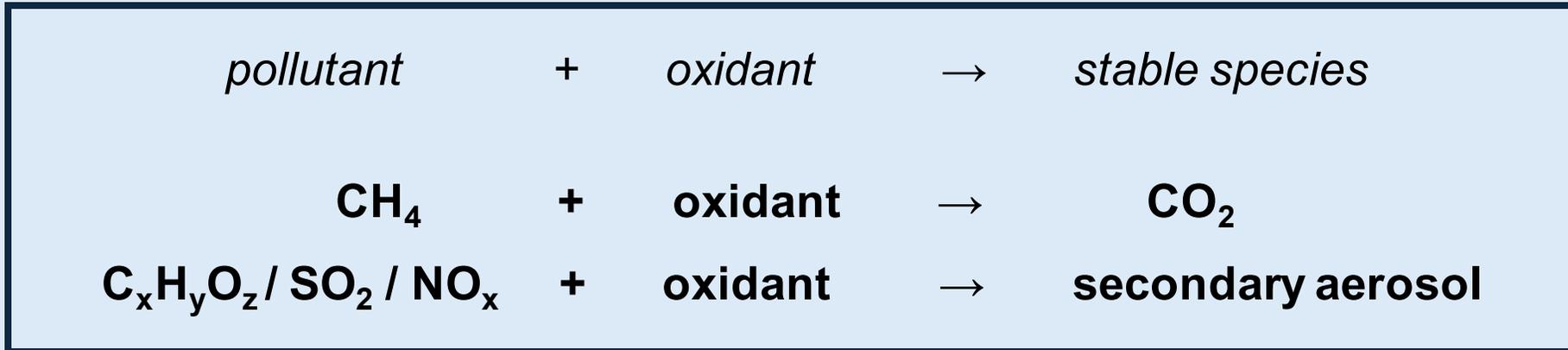


Ice Core and Aerosol Sample Preparation and Nitrate Isotopic Analysis by Electrospray-Orbitrap

Jack Saville & Julien Witwicky, Elsa Gautier, Nicolas Caillon, Joël Savarino



Motivation: Atmospheric Chemistry

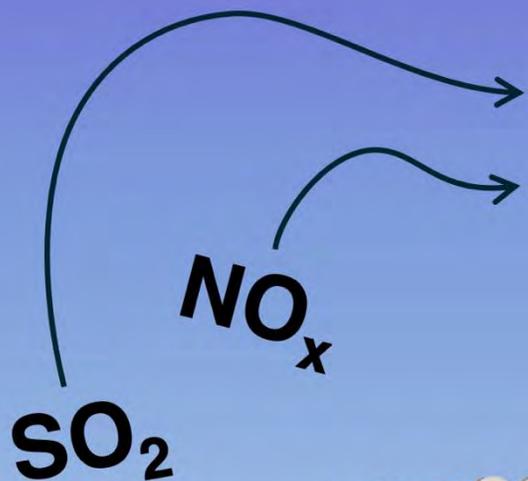


Oxidants [OH], [O₃], [H₂O₂] & [NO₃] control:

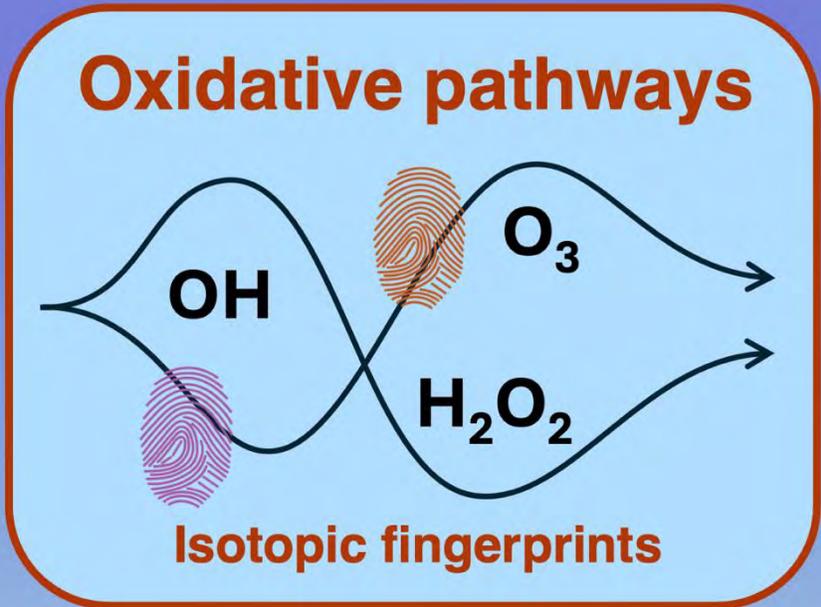
- **pollutant** levels
- tropospheric **ozone** production
- **aerosol** and **cloud** formation
- **methane** lifetime

**Air quality
&
Climate**

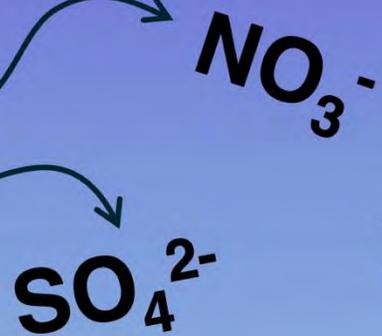
Emissions



Oxidative pathways



Aerosols



Ice core archive



Traditional nitrate isotope method:

Denitrifier IRMS



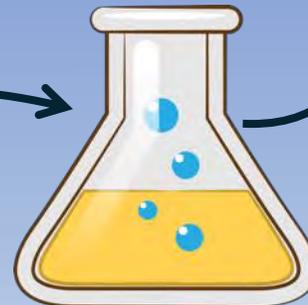
Ice core archive



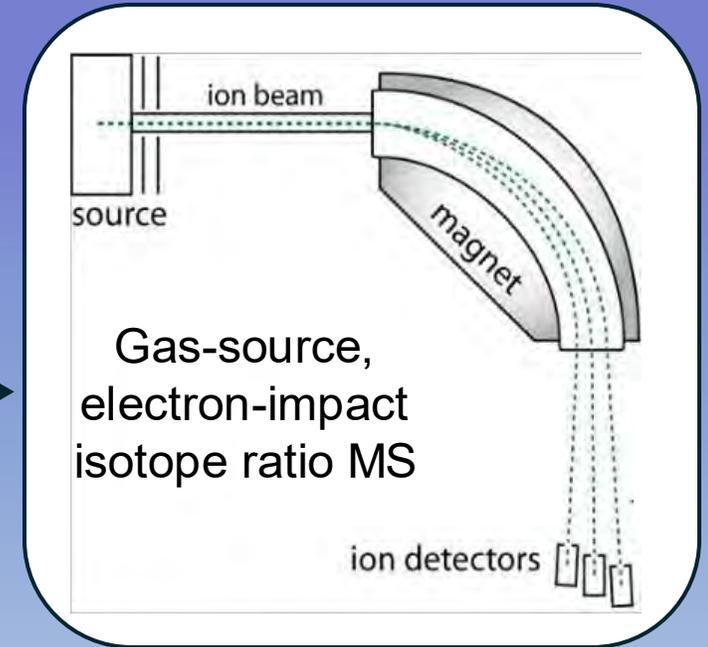
Ice core samples



Concentration and/or purification



Chemical conversion of oxyanions to simple gases



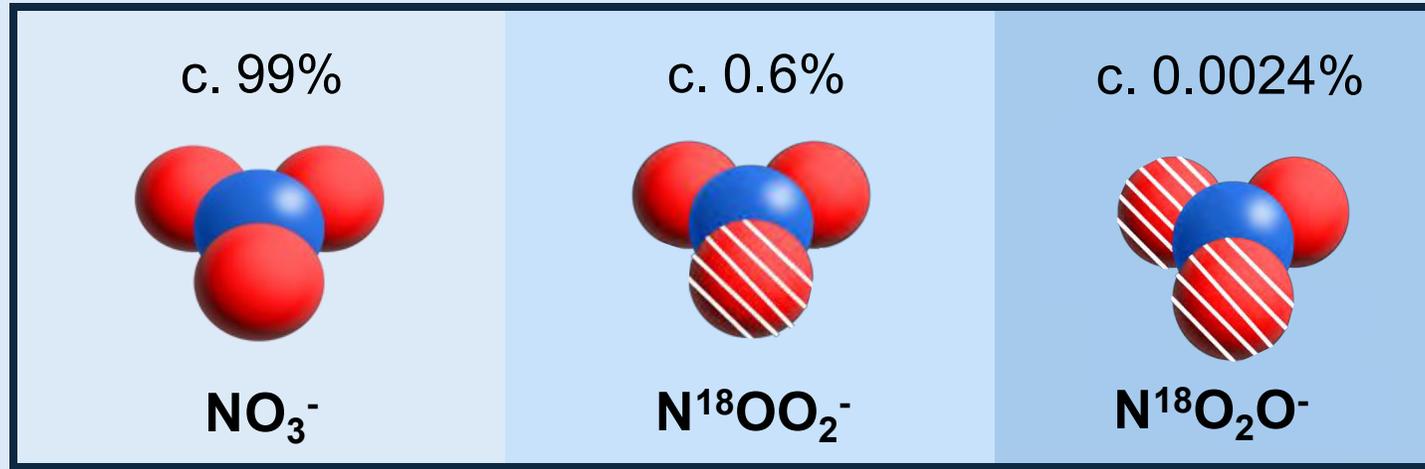
Traditional nitrate isotope method: **Denitrifier IRMS**



Limitations:

1. **Sample size is large.** 100 nmol nitrate ~ 100g alpine ice, 1kg polar ice
2. **O atom recovery is non-quantitative** → accurate nitrate $\delta^{18}\text{O}$ requires careful calibration and reproducible bacteria performance, so can be imprecise
3. **Chemical bonds are broken** → intramolecular information is lost

What are clumped isotopes?



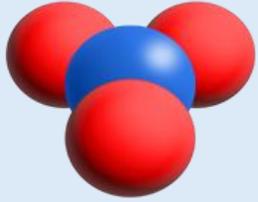
Are isotopes *stochastically distributed* between molecules?

$$R_{1818} = (R_{18})^2$$

Or do **heavy isotopes** prefer to *clump* together in the same molecule?

$$R_{1818} > (R_{18})^2$$

What are clumped isotopes?



$$\Delta^{1518} = \frac{R_{1518}}{R_{15} * R_{18}} - 1$$

$$\Delta^{1818} = \frac{R_{1818}}{R_{18} * R_{18}} - 1$$

$$\Delta^{1718} = \frac{R_{1718}}{R_{17} * R_{18}} - 1$$

Clumped isotopes signatures are:

- imprinted during **molecule-forming reactions**
- dependent on reaction **energetics and kinetics**
 - **insensitive to bulk isotopes**

clumped isotopes

1900 - 2022

English

Case-Insensitive

Smoothing

Biological signatures in clumped isotopes of O₂

2015

Laurence Y. Yeung^{1,2*†}, Jeanine L. Ash^{1*†}, Edward D. Young¹

The abundance of molecules containing more than one rare isotope have been applied

Paleoclimate reconstruction using carbonate clumped isotope thermometry

John M. Eiler*

2011

Division of Geological and Planetary Sciences, California Institute of Technology, Pasadena, CA 91125, USA

Methane clumped isotopes: Progress and potential for a new isotopic tracer

2017



Peter M.J. Douglas^{a,b,*}, Daniel A. Stolper^{a,c}, John M. Eiler^a, Alex L. Sessions^a, Michael Lawson^d, Yanhua Shuai^{a,e}, Andrew Bishop^f, Olaf G. Podlaha^g, Alexandre A. Ferreira^h, Eugenio V. Santos Neto^h, Martin Niemannⁱ, Arne S. Steen^j, Ling Huang^e, Laura Chimiak^a, David L. Valentine^k, Jens Fiebig^l, Andrew J. Luhmann^m, William E. Seyfried Jr.ⁿ, Giuseppe Etiope^{o,p}, Martin Schoell^q, William P. Inskeep^r, Jan

Clumped isotope signatures of nitrous oxide formed by bacterial denitrification

2022

Kristýna Kantnerová^{a,b,*}, Shohei Hattori^{c,d}, Sakae Toyoda^d, Naohiro Yoshida^d, Lukas Emmenegger^a, Stefano M. Bernasconi^b, Joachim Mohn^a

A R T I C L E
0.00000014
0.00000012
0.00000010
0.000000080%
0.000000060%
0.000000040%
0.000000020%
0.000000000%

1900 1910 1920 1930 1940 1950 1960 1970 1980 1990 2000 2010 2020

clumped isotopes

Introducing Electrospray-Orbitrap

A high-sensitivity, liquid-source, soft-ionisation, high-resolution mass spectrometer optimised for accurate-mass identification and quantification of biomolecules

1 10x more productivity, faster analysis by eliminating several sample introduction systems and multiple IRMS systems from the workflow

2 10x less sample preparation, e.g. by avoiding time consuming (and error-inducing) microbial processing, wet chemistry and mineral precipitation

3 Reduced sample sizes due to increased sensitivity compared to classical IRMS approaches

4 Direct measurements of polar analytes in liquid samples without chemical derivatization or conversion

5 Analysis of intact molecules using soft electrospray ionization

6 Simultaneous acquisition of all major and many minor isotopologues by high-resolution accurate mass IRMS

7 Extraction of accurate isotopic information from singly- and multi-substituted isotopologues utilizing methodology for amplifying signals from minor isotopologues

8 Position specific isotope information from functional groups through controlled fragmentation of molecular ions

9 Utilization of proven Dual Inlet principles for determination of accurate isotope ratios of unknown samples relative to a reference

10 Automation of sample introduction by coupling the Orbitrap Exploris Isotope Solutions to Thermo Scientific™ Vanquish™ Neo UHPLC system

thermo scientific

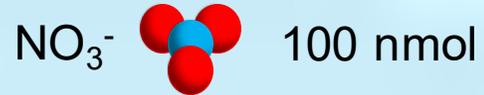
Recently rebranded as an isotopologue-ratio MS

- **Sample demand 10x lower** than IRMS
- **No chemical conversions =** less chances for fractionation
- Intact molecules = **clumped isotopes** are accessible
- **Any soluble molecule** with $m/z > 40$ analysable

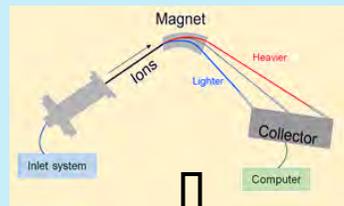
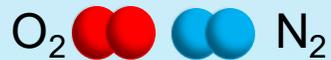
Thermo Scientific

Electrospray-Orbitrap MS instrument

Denitrifier-IRMS



Bacterial conversion



Electrospray-Orbitrap MS



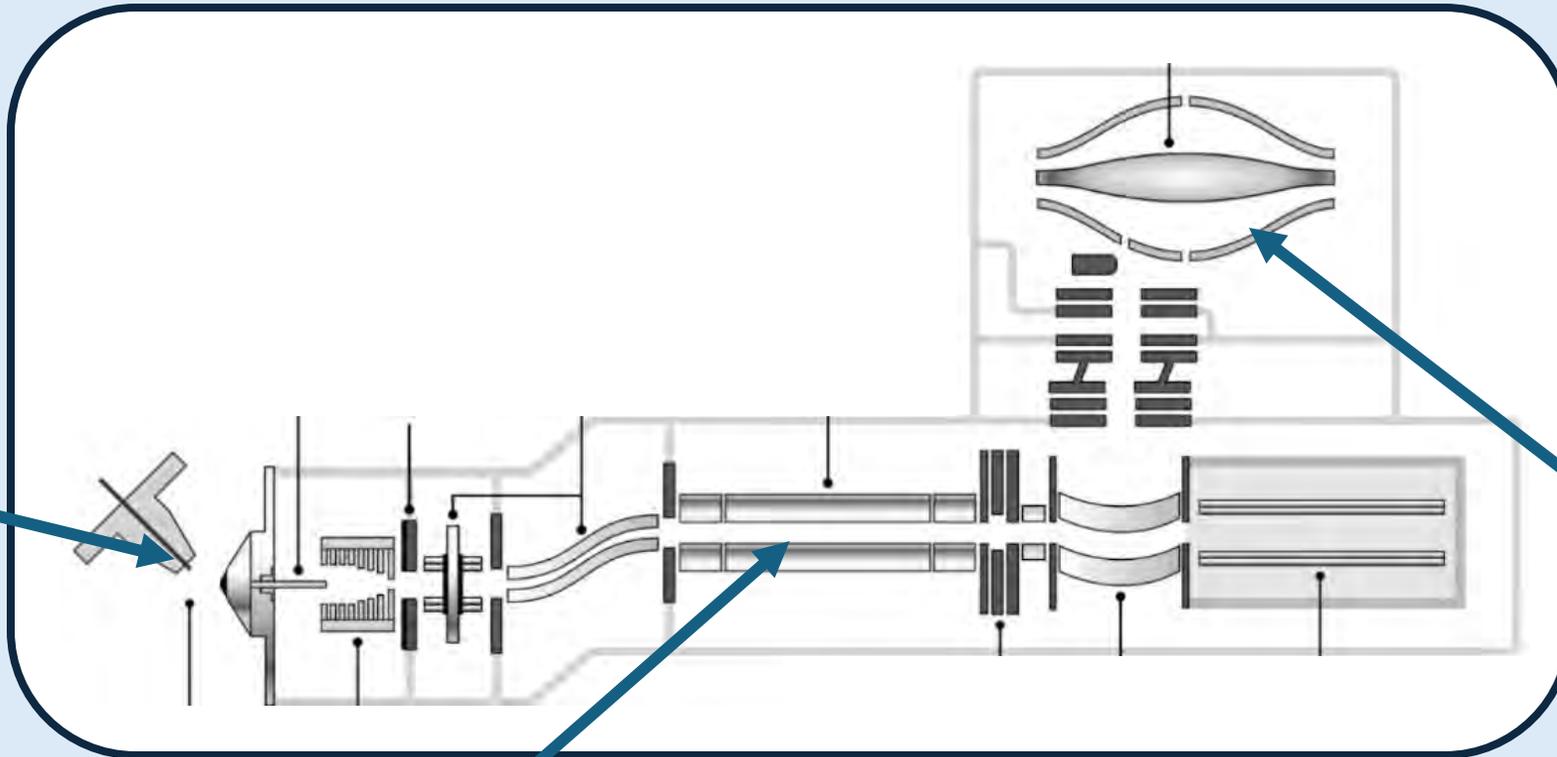
Intact nitrate ions



Electrospray-Orbitrap MS instrument

Pure **methanolic analyte solutions**
50 μM , 4 $\mu\text{L}/\text{min}$
stable delivery

Electrospray ionisation (ESI)
produces
gaseous
molecular ions



Fourier-transform of image current gives
mass spectrum.
Mass resolution \sim
measurement
duration

Orbitrap analysis:
axial ion frequency
 $\omega \propto \sqrt{m/z}$

Quadrupole filter isolates a
mass window for analysis

Successive **fixed-size ion packages** are trapped for analysis.
Package size limited by space-charge effects

Orbitrap Exploris 240, Thermo Fisher; Kantnerová, Kuhlbusch et al. Nature Protocols (2024)

Instrumental constraints

ESI constraints:

Matrix-sensitivity (salts and organics)

Requires **methanolic** samples

Orbitrap constraints:

Package size at least $< 10^6$ ions, or space-charge effects

Scan rate = 30 packages/sec (at 15k resolution)

→ c. **10^7 ions counted per second**

many minutes acquisition required for **1 ‰ precision** $\delta^{15}\text{N}$, $\delta^{18}\text{O}$, $\Delta^{17}\text{O}$,

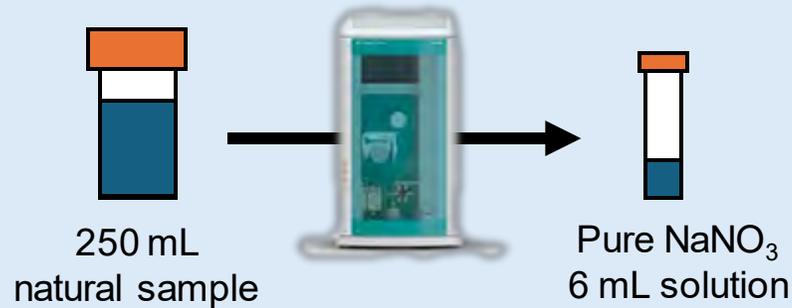
→ LC-MS coupling difficult

Fun bonus constraint!

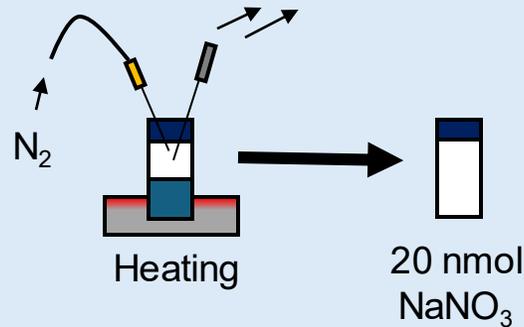
Isotopologue ratio drift affects precision over **> 15 min** timescales

ESI-Orbitrap sample preparation

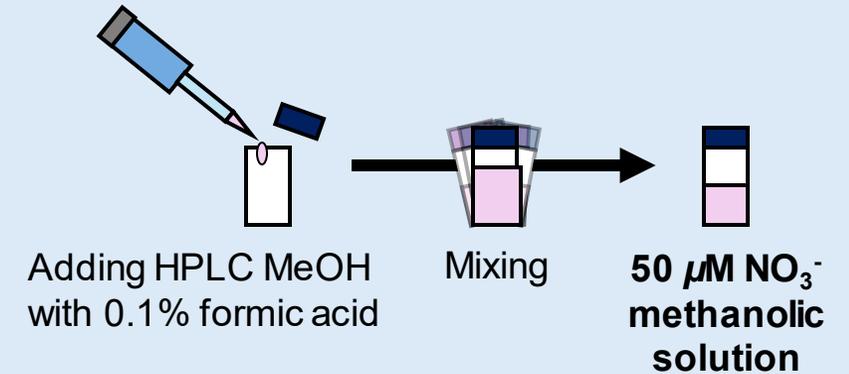
Separation, purification and concentration by IC



Precipitation by heating and N_2 headspace flushing

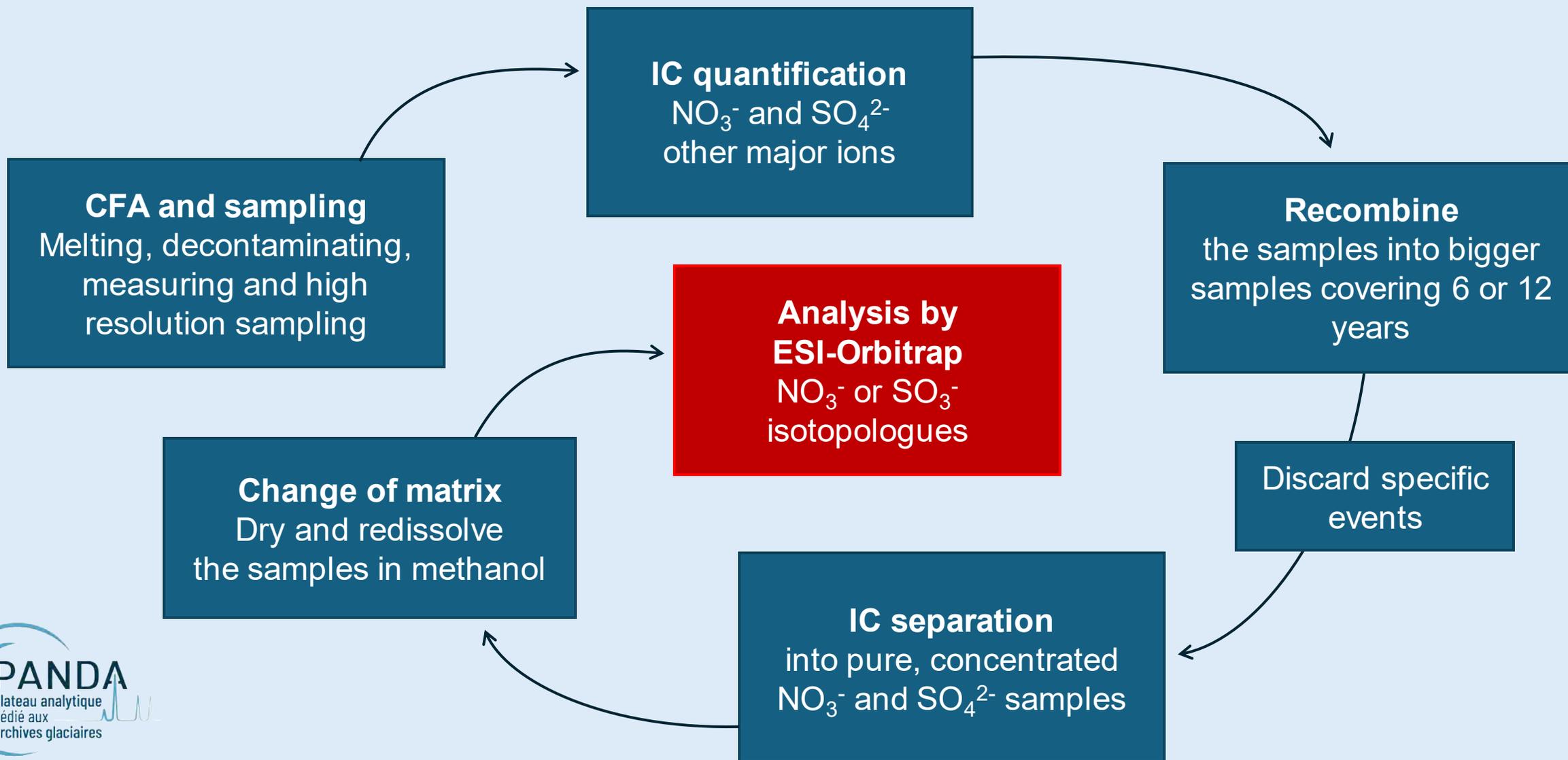


Redissolution in acidified methanol

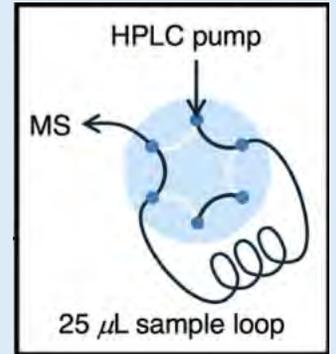


- Aim for simple, consistent matrix
- Principle of identical treatment of samples and calibration standards

Ice core sample preparation



Sample introduction

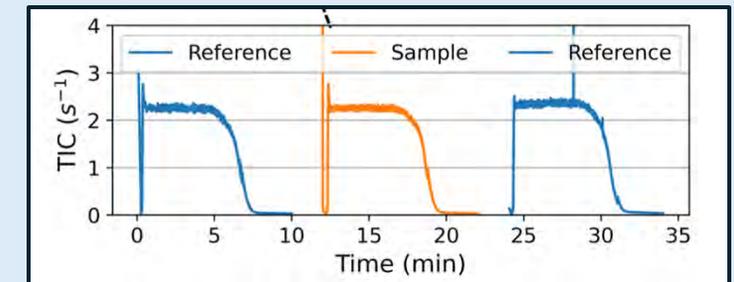


1. Dual inlet via twin syringes

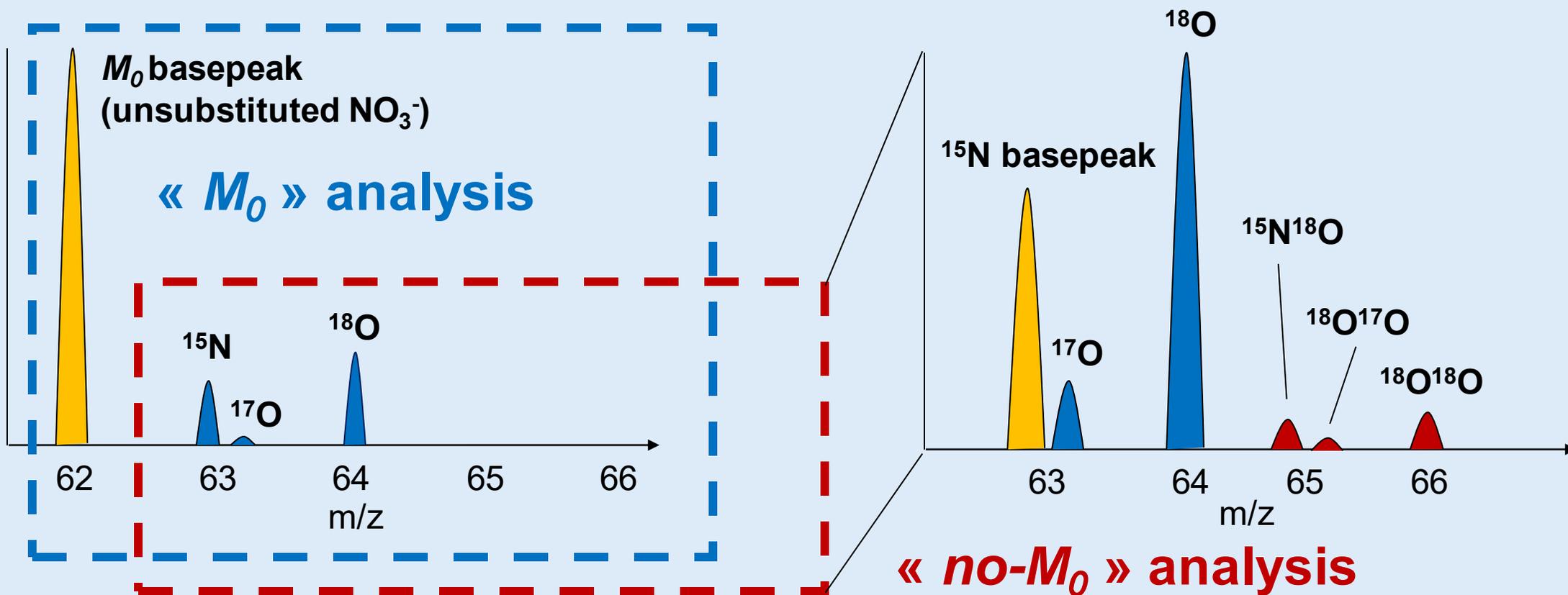
- + Alternating analysis of sample/ref
- + Requires no extra hardware
- + Simplest
- + Time-efficient = less drift = more precise
- 2-point calibration impossible
- Labour-intensive
- No autonomy
- 50 % of sample wasted
- Low reproducibility
- Blanks ~ syringe rinsing dedication

2. In-flow injection via HPLC autosampler

- + Long, autonomous sequences
- + Reproducible injections
- + Many standards and samples in one sequence
- + Controlled memory effect, low blanks
- + More sample-efficient
- Time wasted rinsing sample loop
- HPLC is over-qualified for the job



Isotopologue analysis



Removing the M_0 peak amplifies substituted isotopologue counting and allows quantification of doubly-substituted isotopologues

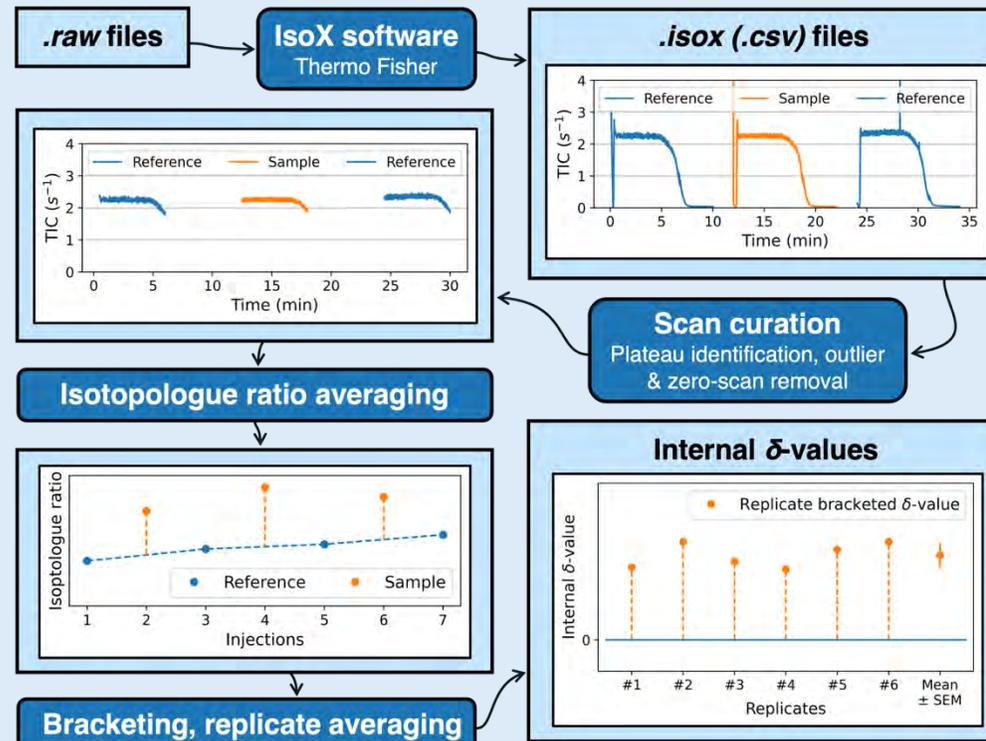
Data acquisition

Bracketed acquisitions

Repeated 6 times
in random order

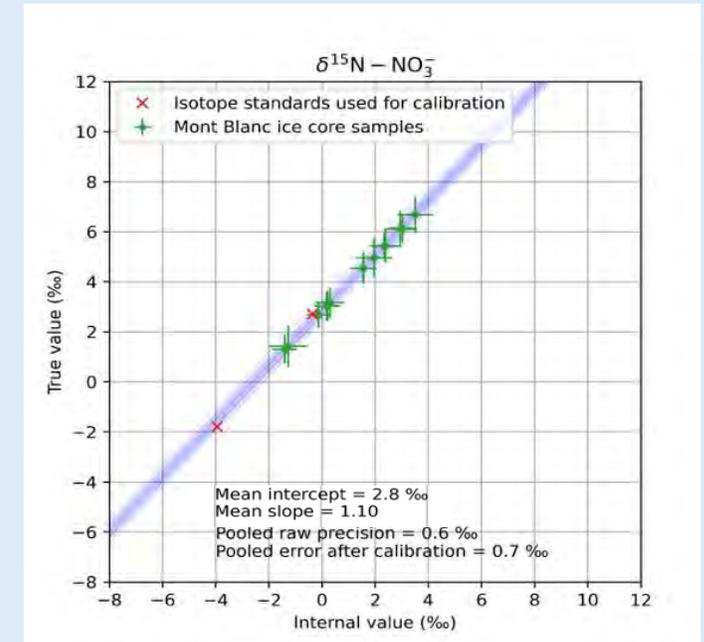
Internal standard	USGS35-ISD
Sample	CdM-28
Internal standard	USGS35-ISD
Sample	CdM-surf1
Internal standard	USGS35-ISD
Calibration standard	USGS32-2
Internal standard	USGS35-ISD
Sample	CdM-20
Internal standard	USGS35-ISD
Calibration standard	USGS35-1
Internal standard	USGS35-ISD
Calibration standard	USGS35-3
Internal standard	USGS35-ISD
Sample	CdM-3
Internal standard	USGS35-ISD
Calibration standard	USGS35-1
Internal standard	USGS35-ISD
Sample	CdM-15
Internal standard	USGS35-ISD

Raw data treatment



Isotopic calibration

Monte-Carlo linear regressions



Standard	$\delta^{15}\text{N}$ (‰)
USGS34	-1.8
USGS35	2.7
USGS32	180.0

Results

This is a preprint and has not been peer reviewed. Data may be preliminary.

WORKING PAPER |  | 12 JANUARY 2026 |  LATEST VERSION    

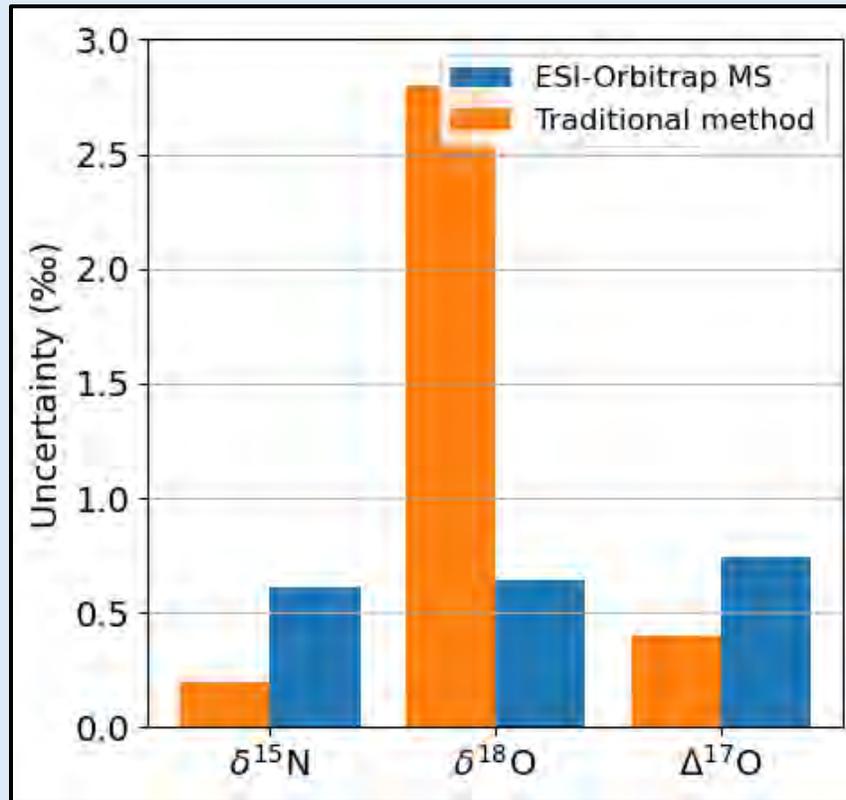
Oxyanion Isotopologue Analysis of Natural Samples by Electrospray-Orbitrap: Method, Validation and Clumped Isotopes

Authors: [Mr. Jack Saville](#)   , [Mr. Julien Witwicky](#), [Dr. Elsa Gautier](#), [Dr. Alexis Lamothe](#), [Mr. Nils Johannes Kuhlbusch](#), [Dr. Becky Alexander](#), [Dr. Andrew Schauer](#), ... [SHOW ALL ...](#) , and [Dr. Joël Savarino](#)  | [Authors Info & Affiliations](#)

<https://doi.org/10.26434/chemrxiv-2026-80tdd> 

 298  281    

Results: nitrate precision



Uncertainty ²

= standard error on mean raw value ²
+ calibration uncertainty ²

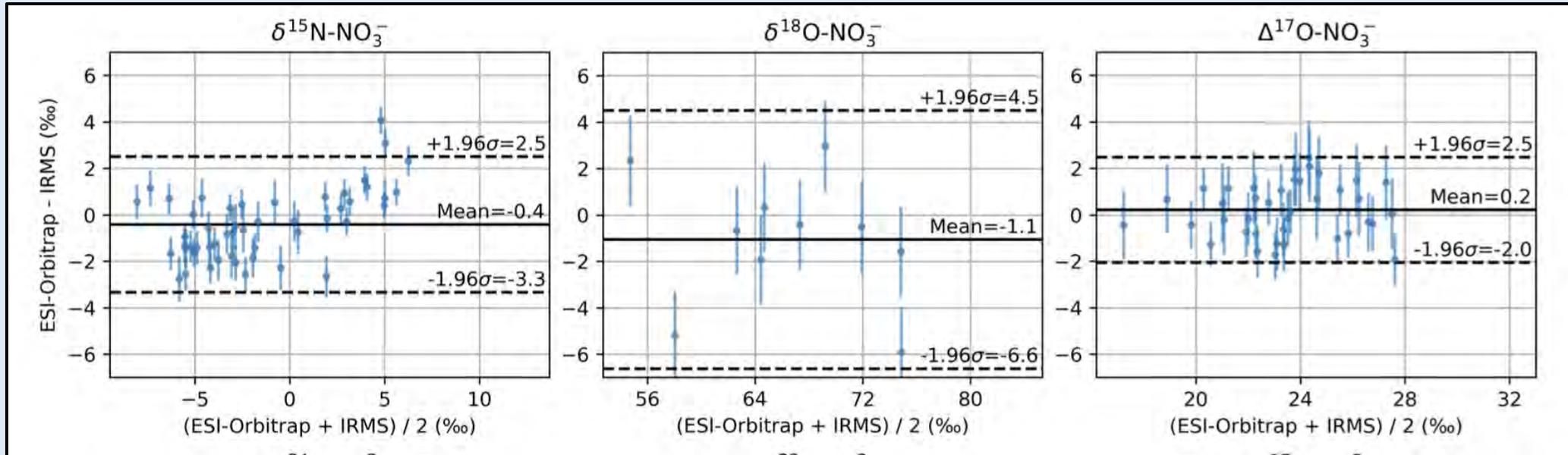
Now we perform even better than this:

$\delta^{15}\text{N} \rightarrow 0.3 \text{ ‰}$

$\delta^{18}\text{O} \rightarrow 0.5 \text{ ‰}$

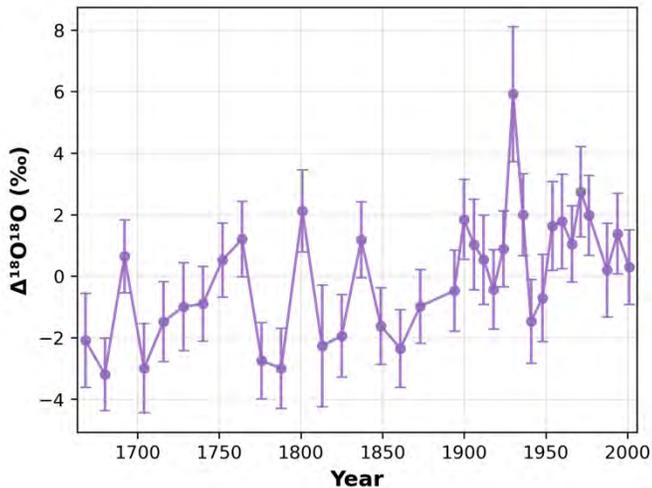
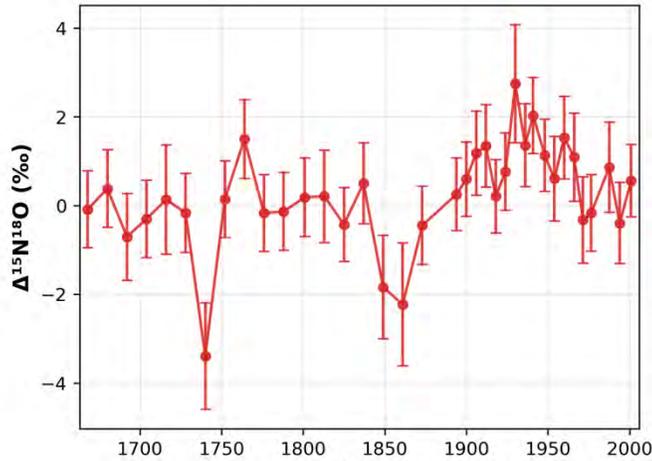
$\Delta^{17}\text{O} \rightarrow 0.5 \text{ ‰}$

Results: nitrate accuracy



	$\delta^{15}\text{N}$	$\delta^{18}\text{O}$	$\Delta^{17}\text{O}$
<i>N</i> ice samples	50	10	39
Mean difference Orbitrap-IRMS (‰)	1.1 ± 0.6	2.1 ± 2.9	0.9 ± 0.9
Mean bias (Orbitrap-IRMS) (‰)	-0.4	-1.1	0.2

Results: nitrate clumped isotopes



Nitrate $\Delta^{15}\text{N}^{18}\text{O}$ and $\Delta^{18}\text{O}^{18}\text{O}$ are measured with **uncertainty 1 – 1.5 ‰** using 20 nmol samples from EGRIP ice core (Greenland)

Measurements are calibrated by assuming intramolecular stochasticity in existing isotope standards

Value are close to zero with variance roughly explainable by measurement uncertainty

→ **Ice core nitrate clumped isotope signals have probably < 1 ‰ amplitude**

Results

- Orbitrap nitrate $\delta^{15}\text{N}$ and $\Delta^{17}\text{O}$ agree with IRMS
- Orbitrap approaching IRMS **precision**
- **Sample size reduced**
100 nmol → 20 nmol
- Nitrate **clumped isotopes** are now measurable

Challenges

- Orbitrap – IRMS nitrate $\delta^{18}\text{O}$ **disagreement**
- **Matrix effects:**
contamination control,
coelutions
- Improve clumped isotope **precision**

Perspectives

- **Nanomole** nitrate samples
= work in clean room
- Nitrate in **new matrices**
(lakes, soils...) require new
purification steps?
- **New analytes** (oxyanions,
secondary organic aerosol,
amino acids, lipids,
metals...)
- **Position-specific** isotope
analysis...

Other analytes

Stable Isotope Analysis of Intact Oxyanions Using Electrospray Quadrupole-Orbitrap Mass Spectrometry

Cajetan Neubauer,* Antoine Crémère, Xingchen T. Wang, Nivedita Thiagarajan, Alex L. Sessions, Jess F. Adkins, Nathan F. Dalleska, Alexandra V. Turchyn, Josephine A. Clegg, Annie Moradian, Michael J. Sweredoski, Spiros D. Garbis, and John M. Eiler

Oxygen Isotope Analyses of Phosphate and Organophosphorus Compounds by Electrospray Ionization Orbitrap Mass Spectrometry

Nora M. Bernet, Cleo Soldini, Timo M. O. Felder, Kristýna Lapčíková, Cajetan Neubauer, Wendy L. Queen, Ralf Kaegi, Federica Tamburini, and Thomas B. Hofstetter*

Simultaneous, High-Precision Measurements of $\delta^2\text{H}$ and $\delta^{13}\text{C}$ in Nanomole Quantities of Acetate Using Electrospray Ionization-Quadrupole-Orbitrap Mass Spectrometry

Elliott P. Mueller,* Alex L. Sessions, Peter E. Sauer, Gabriella M. Weiss, and John M. Eiler

Triple Oxygen Isotope Analysis of Methanesulfonate Using the SO_3^- Fragment in ESI-Orbitrap-MS

Yihang Hong, Longchen Zhu, Daniel R. Crocker, Tengyu Liu, Zhenfei Wang, Zhao Wei, Chen Yu, Yu Wei, Hao Yan, David T. Johnston, Issaku E. Kohl, Yongbo Peng, Andreas Hilkert, Cajetan Neubauer, and Shohei Hattori*

Metal Ion Isotope Ratio Using ESI-Orbitrap HRMS: Proof of Concept and Initial Performance Evaluation for Lead Isotopic Ratios

Gianluca Roncoroni, Davide Spanu, Gilberto Binda, and Damiano Monticelli*

Sub-10 Nanomole Perchlorate $\delta^{37}\text{Cl}$, $\delta^{18}\text{O}$, and $\Delta^{17}\text{O}$ Measurements by ESI-Orbitrap-MS

Longchen Zhu, Yihang Hong, Shohei Hattori, Zhenfei Wang, Zhao Wei, Yongbo Peng, Nils Johannes Kuhlbusch, and Huiming Bao*

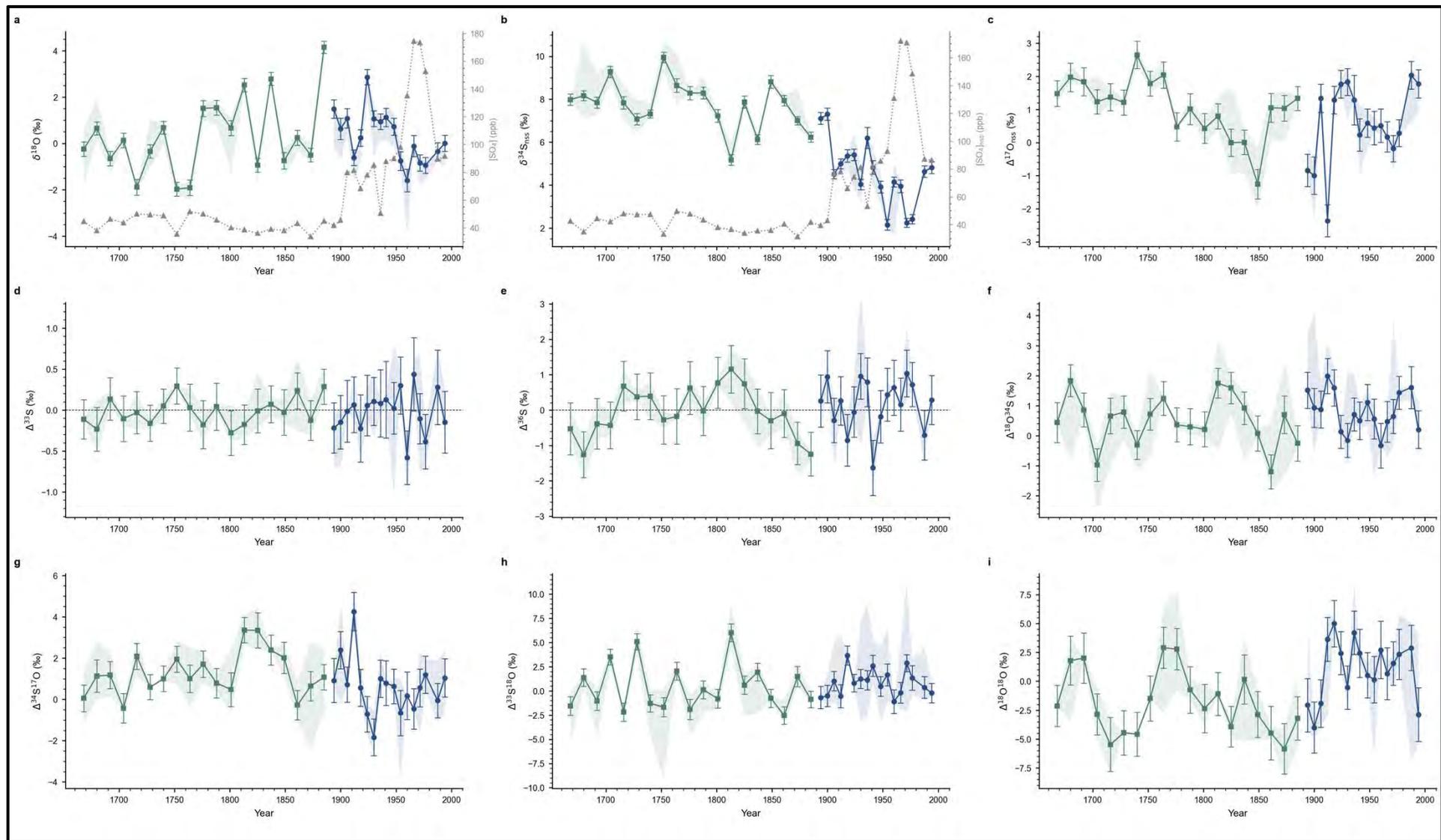
Advancing Stable Isotope Analysis with Orbitrap-MS for Fatty Acid Methyl Esters and Complex Lipid Matrices

Gabriel F. dos Santos,* Giovanni B. Bevilaqua, Alexis Gilbert, Hugo G. Machado, Maxime Julien, Gesiane S. Lima, Nerilson M. Lima, Júlio C. O. Ribeiro, Alexandre A. Ferreira, Ygor S. Rocha, and Boniek Gontijo*

Stable Carbon Isotope Analysis of PFOA and PFOS Using Orbitrap Mass Spectrometry

Hiroto Kawashima,* Tomoha Iezumi, Momoka Suto, and Sachi Taniyasu

Sulphate isotopologue analysis



Nitrate oxygen isotopes

